**Solid State Nanopores**

# **Nanopore Formation via Tip-Controlled Local Breakdown Using an Atomic Force Microscope**

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**The dielectric breakdown approach for forming nanopores has greatly accelerated the pace of research in solid-state nanopore sensing, enabling inexpensive formation of nanopores via a bench top setup. Here the potential of tip-controlled local breakdown (TCLB) to fabricate pores 100× faster, with high scalability and nanometer positioning precision using an atomic force microscope (AFM) is demonstrated. A conductive AFM tip is brought into contact with a silicon nitride membrane positioned above an electrolyte reservoir. Application of a voltage pulse at the tip leads to the formation of a single nanoscale pore. Pores are formed precisely at the tip position with a complete suppression of multiple pore formation. In addition, the approach greatly accelerates the electric breakdown process, leading to an average pore fabrication time on the order of 10 ms, at least two orders of magnitude shorter than achieved by classic dielectric breakdown approaches. With this fast pore writing speed over 300 pores can be fabricated in half an hour on the same membrane.**

## **1. Introduction**

Following successful demonstration of nanopore sequencing via engineered protein pores,<sup>[1]</sup> the next research frontier in nanopore physics is the development of solid-state nanopore devices with sequencing or diagnostic capability.<sup>[2]</sup> Solid-state pores are mechanically more robust, admit of cheaper, more scalable fabrication, have greater compatibility with complementary metal-oxide-semiconductor (CMOS) semiconductor technology,

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possess enhanced micro/nanofluidic integration potential $[3]$  and could potentially increase sensing resolution.[2] Yet, despite the great interest in solid-state pore devices, approaches for fabricating solid-state pores, especially with diameters below 10 nm, are limited, with the main challenge being a lack of scalable processes permitting integration of single solid-state pores with other nanoscale elements required for solid-sate sequencing schemes, such as transverse nanoelectrodes,[4,5] surface plasmonic structures, $[6-10]$  and micro/nanochannels.[11–14] The main pore production approaches, such as milling via electron beams in a transmission electron microscope  $(TEM)^{[15]}$  and focused-ion beam  $(FIB)$ ,<sup>[16–18]</sup> use high energy beam etching of substrate material. While these tech-

niques can produce sub 10 nm pores with nm positioning precision, they require expensive tools and lack scalability.

In 2014 Kwok et al.<sup>[19,20]</sup> showed that by directly applying a voltage across an insulating membrane in electrolyte solution, they could form single nanopores down to 2 nm in size. The applied voltage induces a high electric field across the thin membrane, so strong that it can induce dielectric breakdown, leading to pore formation. The dielectric breakdown method is fast, inexpensive and potentially highly scalable, yet it has a critical disadvantage: the pore position is random. When a high transmembrane voltage is applied electric breakdown occurs at a "weak" location on the insulating membrane, a position determined randomly by the intrinsic inhomogeneity of the nitride film. As the pore can form anywhere on the membrane upon voltage application, the breakdown technique cannot form pores at precisely determined positions; creating multiple pores with welldefined spacing is likewise unfeasible. This is a very problematic limitation, particularly given that many solid-state sensing and sequencing schemes requiring precise pore positioning (e.g., between transverse electrodes, $[4,5]$  carbon nanotubes, $[21]$  graphene nanoribbon,<sup>[22]</sup> or within a micro/nanofluidic channel<sup>[11-13]</sup>). Multiple closely spaced pores show promise for translocation control.[12,13,23] Critically, the breakdown approach may also inadvertently produce more than one nanopore over the membrane area,<sup>[24–27]</sup> leading to a drastic loss of signal-to-noise and inability to resolve single-molecule translocation events. A recent variation of the breakdown approach uses a pipette tip to control voltage application,[28] increasing pore positioning precision to the micron scale (the pipette tip opening diameter is 2 μm),





**Figure 1.** Nanopore fabrication via tip-controlled local breakdown (TCLB). A 3D schematic of the experimental setup depicting an AFM cantilever with a conductive tip positioned over a silicon nitride membrane. Application of a voltage pulse leads to formation of a nanopore at the tip position. Nanopore arrays can be readily formed via control of the AFM tip location, with in situ current measurement at each pore verifying successful pore fabrication at that location. Note that our setup requires only one side of the membrane to be in contact with electrolyte, while the other side of the membrane is exposed to air.

but nanometer positioning precision is in fact required for many solid-state sequencing schemes, due to the small size of sensing elements required to interface with the pores. In addition, the pipette-tip approach does not prevent the potential formation of multiple pores over the still large (micrometer scale) region of voltage application.

We have developed a new approach for forming solid-state pores that combines the positioning advantages of particle beam milling and the simplicity/low-cost of the electric breakdown approach with the powerful imaging capabilities of atomic force microscopy (AFM). In our approach, which we call tip-controlled local breakdown (TCLB), a conductive AFM tip is brought into contact with a silicon nitride membrane and used to apply a local voltage to the membrane (**Figure 1**). The local voltage induces electric breakdown at a position on the membrane determined by the AFM tip, forming a nanopore at that location, which we demonstrate via *I*–*V* measurement, TEM characterization, and single-molecule translocation. TCLB has several advantages. First, in TCLB, the nanoscale curvature of the AFM tip ( $r \approx 10$  nm) localizes the electric field to a truly nanoscale region, eliminating the possibility of forming undesirable additional nanopores on the membrane as well as preventing the pore-free region of the membrane from being damaged by high electric fields. Secondly, TCLB can form pores with a spatial precision determined by the nanoscale positioning capability of the AFM instrument (an improvement in spatial precision from micro to nanoscale). Third, TCLB drastically shortens the fabrication time of a single nanopore from on order of seconds to on order of 10 ms (an improvement of at least 2 orders of magnitude). Fast pore fabrication implies that arrays can be written with extremely high throughput (over  $\approx$ 100 pores in half an hour, compared to  $\approx$ 100 in a day<sup>[28]</sup>). Fourthly, as TCLB is AFM based, it can harness the topographic, chemical and electrostatic scanning modalities of an AFM to image the membrane before and after pore formation, enabling precise alignment of pores to existing features. The scanning capabilities of the AFM tool can be used to automate fabrication of arrays of precisely positioned pores, with the

successful fabrication of each pore automatically verified by current measurement at the tip following voltage application. The precise control of the contact force, made possible by AFM, is essential for establishing the reliable contact between the tip and the membrane. As AFM are benchtop tools that operate in ambient conditions (e.g., at atmospheric pressure and normal indoor humidity) they are inherently low-cost and can be readily scaled. The ability to work in ambient conditions implies that the approach is compatible with materials possessing which require sensitive chemical functionalization (e.g., that might be damaged by vacuum conditions used in FIB and TEM). Finally, while classic dielectric breakdown requires that both sides of the membrane be in contact with aqueous electrolyte reservoirs, our approach requires that only one side of the membrane be in contact with a liquid reservoir, considerably easing the scaling of our method and the speed of nanopore formation, as the AFM scanning takes place in a dry environment.

## **2. Results**

### **2.1. Nanopore Fabrication**

The schematic of the experimental setup is illustrated in Figure 1. Using a bench-top AFM setup operated in ambient laboratory conditions, a conductive AFM tip is brought into contact with a thin silicon nitride membrane sitting on top of an electrolyte reservoir. The conductive AFM tip is positioned a distance of ≈ 100 μm from the membrane (**Figure 2**a). To initiate pore fabrication, the tip approaches the membrane at a speed of  $\approx$ 5 µm s<sup>-1</sup> until it engages with the surface (Figure 2b). A small loading force (typically in the order of 1 nN) is applied to the tip in order to minimize contact resistance between the tip and the membrane. This force is set sufficiently small to avoid tip-induced mechanical damage to the membrane. To initiate the breakdown process, the tip is positioned at the desired location in the scanning region and a single rectangular pulse is applied (Figure 2c). The pulse has an amplitude of V<sub>pulse</sub>, and a duration of  $t_{\text{pulse}}$ . The applied voltage pulse initiates the breakdown process and creates a nanoscale pore on the membrane, located at the tip location. After nanopore formation, the tip is retracted from the membrane (Figure 2d). A representative breakdown event is shown in Figure 2e–g. A voltage pulse of  $V_{\text{pulse}} = 24$  V,  $t_{\text{pulse}} = 100$  ms is applied (Figure 2e). After voltage application, the current increases to  $\approx 50$  pA and remains roughly constant (Figure 2f, inset). After a time delay of  $t_{BD}$  = 36.2 ms (Figure 2f), the current increases sharply to a few nA, indicating successful breakdown and nanopore formation. If the pores are large, successful nanopore fabrication at the tip location can additionally be confirmed by a subsequent topographic AFM scan (Figure 2h,i). When the nanopore diameter is smaller or comparable  $(d \leq 10 \text{ nm})$  to the tip radius of curvature, the nanopore may not be observed in the AFM scan.

We have developed a custom script enabling automatic control of the pore fabrication process. Using this script we can readily create pore arrays, iterating the single-pore formation process over a  $5 \times 5$  grid with the pores spaced evenly by 500 nm. Using the same tip, we have successfully fabricated over 300 nanopores on the same membrane, demonstrating the

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 $\mathbf b$ d a  $\mathsf C$ Tic SiNx Electrolyte Approach **Breakdown** Retraction Current (nA) Voltage (V) 20 Voltage l<sub>oulse</sub>  $\mathsf{V}_{\mathsf{pulse}}$  $10$  $\Omega$  $\rm t_{BD}$  $10$  ${\rm t_{BD}}$ Current 5 H RC decay Voltage Pore Voltage applied creation Voltage applied cut-off  $\mathbf 0$  $0.0$  $0.2$  $0.6$  $0.8$  $1.0$  $0.4$ Pore creation Time (s) 41.3 nm  $R_a = 0.66$  nm I nanopore 35.0 لمما Height (nm)  $30.0$ Line 1 25.0 150 300 450 600  $30$ 20.0 15.0  $1!$  $10.0$ Line 2 400 nm  $10 \text{ nm}$ 0  $0<sub>0</sub>$  $\frac{1}{150}$ 450  $\overline{600}$ 300 X position (nm)

**Figure 2.** Fabrication process of a single nanopore by conductive tip induced local electric breakdown. a) Schematic showing the conductive AFM tip located above a thin silicon nitride membrane. The bottom side of the membrane is in contact with electrolyte. b) To minimize contact resistance between the tip and membrane, the tip is pressed against the membrane in contact mode. c) A voltage pulse is applied across the membrane through the tip, initiating the breakdown process, resulting in the formation of a single nanopore. d) Tip is retracted from the membrane once a nanopore is formed. The voltage pulse e) and the current across the membrane f) during a typical nanopore fabrication event. The membrane thickness is 20 nm, the pulse height  $V_{\text{pulse}} = 24$  V, the pulse width  $t_{\text{pulse}} = 100$  ms and the tip radius is  $10 \pm 5$  nm. g) TEM image of a 9.2 nm pore corresponding to the current and voltage trace shown in (e) and (f). h) AFM scan of a larger sized single nanopore fabricated on silicon nitride membrane using TCLB with accompanying topographic scans of bare membrane (i-red, surface roughness  $R_a = 0.66$  nm) and across the pore (i-blue). Note that small nanopores  $(d \leq 10 \text{ nm})$  may not show up on an AFM scan as the tip radius is too large to resolve the small pores.

scalability of our TCLB technique (see Section S2 in the Supporting Information for more information).

#### **2.2. Probing the Breakdown Threshold**

Our automated pore fabrication protocol enables efficient varying of process parameters to optimize pore fabrication.

In particular, we vary the pulse amplitude across the nanopore array to probe the threshold at which membrane breakdown occurs. A pulse train of five subsets, with each set containing five rectangular pulses of fixed duration (100 ms) but increasing amplitude (11–15 V, with an increment of 1 V), are applied across the membrane (**Figure 3**a, blue trace). Each pulse is applied to a different location on the membrane. The detected current is shown in Figure 3b (trace in red).



**Figure 3.** Automatic probing of pore fabrication conditions. a) The voltage pulse train applied to different membrane locations and b) the resulting current. Magnified view of voltage pulse c) and resulting current d) that does not correspond to pore fabrication. Magnified view of voltage pulse e) and resulting current f) that does correspond to pore fabrication. g) Pore formation conditions across the 5×5 array. Location A1 corresponds to (c) and (d); location D4 corresponds to (e) and (f).

The locations are arrayed spatially in a  $5 \times 5$  square grid, with the pulse location in the array given by Figure 3g. The fabrication process starts from location A1 and ends at location E5, rastering in the *y* direction (Figure 3g, A1→A5, B1→B5, C1→C5, D1→D5, E1→E5). The spacing between each fabrication site is 500 nm. Spikes in the detected current, which occur for pulse amplitudes greater than 13 V, indicate successful electric breakdown. At  $V_{\text{pulse}} = 14$  V, 2 out of 5 attempts induce breakdown. A further increase of the voltage to 15 V leads to a 100% breakdown probability (5 out of 5). Magnified view of no-breakdown and successful breakdown events are shown in Figure 3c–f corresponding to location A1 ( $V_{\text{pulse}}$  = 11 V) and D4 ( $V_{\text{pulse}} = 14$  V).

#### **2.3. TEM Characterization**

TEM microscopy allows for a detailed characterization of the nanopores made by TCLB. **Figure 4** shows three TEM micrographs of nanopore arrays. In agreement with our AFM settings (Figure 3g), nanopores are spaced evenly by 500 nm in an array format. Figure 4a,b shows a  $3 \times 3$  nanopore array fabricated using  $V_{pulse} = 15$  V,  $t_{pulse} = 100$  ms. Figure 4c,d shows two nanopore arrays made on a new membrane (same thickness) with a new tip (same radius of curvature) under exactly the same fabrication conditions ( $V_{\text{pulse}} = 15$  V,  $t_{\text{pulse}} = 100 \text{ ms}.$ 

#### **2.4. Pore Formation Mechanism**

#### *2.4.1. Weibull Versus Log-Normal*

Nanopore fabrication time (time-to-breakdown,  $t_{BD}$ ) can provide insight into the pore formation mechanism. Nanopores fabricated via classic dielectric breakdown have a time-tobreakdown following a Weibull probability distribution.[28–30] The Weibull distribution is used extensively to model the timeto-failure of semiconductor devices.[31,32] The Weibull distribution arises from the "weakest-link" nature of typical dielectric breakdown process, where breakdown happens at the weakest position over a large membrane area. The nanopore fabrication time is dominated by the time to make a pore at this weakest position.

In contrast, we find that our time-to-breakdown distribution, obtained from forming over 300 nanopores using our automatic





**Figure 4.** TEM characterization of nanopore arrays. a) TEM micrograph of a nanopore array containing 9 nanopores. Nanopores are located at the center of the dashed circles. The pore-to-pore spacing is ≈500 nm. b) Zoomed-in TEM micrograph of a nanopore with an opening diameter of 4.1 nm. c,d) TEM micrograph of nanopore arrays fabricated on a different membrane from (a). Insets showing magnified micrographs of different nanopores with diameter close to or under 5 nm. Fabrication condition:  $V_{\text{pulse}} = 15$  V,  $t_{\text{pulse}} = 100$  ms, membrane thickness  $l = 12-14$  nm, tip radius  $r = 10 \pm 5$  nm. Additional examples of nanopore arrays are shown in Figure S3 in the Supporting Information.

process, yields better agreement with a log-normal probability distribution. **Figure 5** shows the cumulative distribution of timeto-breakdown plotted with a log-normal scaling. In this form, data distributed according to a log-normal distribution follows a straight line. Our time-to-breakdown results, linearized by this rescaling, are thus clearly consistent with a log-normal distribution. In Figure S4 (Supporting Information), we plot the same results rescaled appropriately for a Weibull, and it is apparent that the Weibull is not as good a description. See Section S4 in the Supporting Information for more detail on log-normal, Weibull distribution and appropriate rescalings (probability plot forms).

The better agreement with a log-normal suggests that the physical mechanism of pore-formation is different using TCLB than classic breakdown. Under tip control, the membrane location where dielectric breakdown occurs is controlled by the tip position, and is thus highly defined rather than random. In this case the statistics of membrane breakdown is no longer a weakest link problem (i.e., determined by the time to breakdown of some randomly located "weak-point"), but instead is determined by the degradation of a "typical" location on the membrane reflecting average film properties. Theoretical and experimental work demonstrate that the overall time-scale of a degradation process that arises from the multiplicative action of many small degradation steps (regardless of physical mechanism) can be modelled via a log-normal distribution.<sup>[33-36]</sup> Possible degradation mechanisms for our pore-formation process include electromigration, diffusion and corrosion.[37]

#### *2.4.2. Voltage Dependence of Time-to-Breakdown*

In **Figure** 6a we show the mean time-to-breakdown  $(\langle t_{BD} \rangle)$ versus voltage on a semilog scale. The mean time-to-breakdown decreases exponentially with voltage. This behavior





**Figure 5.** Log-normal probability plot of time-to-breakdown  $(t_{BD})$  for a total of 308 nanopores under different pulse voltages. a) Cumulative distribution of  $t_{BD}$  presented with a log-normal rescaling under following conditions:  $V_{pulse} = 15$  V,  $t_{pulse} = 100$  ms, membrane thickness  $l = 12-14$  nm. The average nanopore fabrication time is  $\langle t_{\text{BD}} \rangle = 20.9 \pm 1.4$  ms. b–d) Cumulative distributions of  $t_{\text{BD}}$  with  $V_{\text{pulse}} = 16, 17, 18$  V, respectively. The dashed lines give the best fit to a log-normal distribution. All experiments are performed with the same tip on one membrane. Tip radius of curvature: ≈10 nm. Membrane thickness: 12–14 nm. Window size:  $50 \times 50 \mu m^2$ . (See Section S4 in the Supporting Information for more details regarding log-normal distribution, Weibull distribution, and probability plots.)

is predicted by the *E*-model of time dependent dielectric breakdown (TDDB),<sup>[38]</sup> which predicts that the mean timeto-breakdown should depend exponentially on the local electric field (proportional to applied voltage at the tip). The *E*-model arises fundamentally from a thermochemical<sup>[38,39]</sup> rather than a direct tunneling mechanism (Fowler-Nordheim tunneling).[40] In thermochemical breakdown, high voltage across the dielectric material induces strong dipolar coupling of local electric field with intrinsic defects in the dielectric. Weak bonding states can be thermally broken due to this strong dipole-field coupling, which in turn serves to lower the activation energy required for thermal bond-breakage and accelerates the degradation process, resulting in a final dielectric breakdown.[38,39]

We have also investigated whether we can use tip-controlled breakdown to produce pores in thicker (20 nm) silicon nitride membranes. We are able to form pores with a high probability but with a corresponding increase in the required voltage, as demonstrated by Figure 6b. The mean time-to-breakdown as a function of voltage in the thicker membranes also follow the *E*-model (Figure 6c).

In Figure 6d we compare the average time-to-breakdown for our tip-controlled approach versus classical dielectric breakdown. We find that our approach gives pore formation times two orders of magnitude lower than classical breakdown, by comparison with a wide-range of experimental studies[8,19,27–30,41–43] exploring classical breakdown for different film thickness (10–30 nm, 75 nm), pH (2–13.5), and voltage (1–24 V).

#### **2.5. Single Molecule DNA Detection**

Lastly, we show nanopores produced using our tip-controlled approach can be used for single molecule detection. **Figure 7** shows results for 100 bp ladder DNA (100–2000 bp) translocating through a 9.9 nm pore ( $V_{\text{pulse}} = 20$  V,  $t_{\text{pulse}} = 150$  ms, membrane thickness 10 nm, tip radius  $r = 10 \pm 5$  nm). To perform single molecule detection, the chip is transferred to a fluidic cell with DNA containing 1 <sup>m</sup> KCl buffer added to the *cis* chamber and DNA-free buffer added to the *trans* chamber. A potential drop of 200 mV is applied across the nanopore, so that DNA molecule are pulled from *cis* to *trans* through the pore. Figure 7a–b shows typical signatures of ionic blockades induced by translocating DNA, composed of a mixture of single and multilevel events. A histogram of current blockades, including 587 translocation events measured by the same nanopore, is shown in Figure 7d. Prior to



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Figure 6. a) Semilog plot of the mean breakdown time ( $\langle t_{\rm BD} \rangle$ ) versus voltage for 12–14 nm thick silicon nitride membrane with an exponential fit. b) Breakdown probability versus voltage for 12–14 and 20 nm thick nitride. c) Semilog plot of the mean breakdown time versus voltage for a 20 nm thick silicon nitride membrane. d) Comparison of average nanopore fabrication time of this work versus range of studies exploring classical breakdown.[8,19,27–30,41,42] Note that if the average fabrication time is not given or cannot be estimated from the reference, a range is then plotted for comparison (see Section S5 more details in the Supporting Information).

performing this DNA translocation experiment, an *I*–*V* trace was obtained to characterize pore size (Figure 7e), which yielded a nanopore resistance of 23.0 MΩ. This strong linearity between current and applied voltage demonstrates that our TCLB fabricated nanopore has an outstanding Ohmic performance. Using a membrane thickness *l* = 10 nm and an electrolyte conductivity  $\sigma = 10$  S m<sup>-1</sup>, according to the pore conductance model<sup>[44]</sup> the estimated effective pore diameter is 9.9 nm.

### **3. Discussion and Conclusion**

In summary, we show that tip-controlled local breakdown can be used to produce pores with nm positioning precision (determined by AFM tip), high scalability (hundreds of pores over a single membrane) and fast formation (100× faster than classic breakdown) using a bench-top tool. These capabilities will greatly accelerate the field of solid-state nanopore research. In particular, the nm positioning is crucial for wide-range sensing and sequencing applications where there is a need to interface nanopores with additional nanoscale elements. Sequencing approaches based on tunneling require positioning a pore between two electrodes.[4,5] Plasmonic devices with interfaced pores require positioning pores at the optimal distance (10–20 nm) from nanoantennas in order to maximize plasmonic coupling.<sup>[6-10]</sup> In devices utilizing nanofluidic confinement (e.g., nanochannels, nanocavities) pores need to be aligned with etched sub 100 nm features.<sup>[11-13,45,46]</sup> In addition to producing pores, our AFM based approach can exploit multiple scanning modalities (topographic, chemical, electrostatic) to map the device prior to pore production and so align pores precisely to existing features.

The ability to control the nanopore diameter is also an important criterion of a pore fabrication technology. In order to control the nanopore diameter via TCLB, we have studied the dependence of the nanopore diameter as a function of pulse width ( $t_{\text{pulse}}$ ) and tip loading force (measured using deflection voltage). Empirically, we found that longer pulse width and higher tip loading force leads to the formation of a larger diameter nanopore (see Section S7 in the Supporting Information). We believe the final nanopore diameter can be adjusted via careful tuning of either of these two parameters (or a combination of both). Alternatively, as reported in previous studies, [28,47] application of voltage pulses across the nanopore after the initial pore formation can enlarge the nanopore. Such a strategy might also be used together with TCLB.

TCLB can be integrated into an automated wafer-scale AFM system, ensuring nm alignment of each pore with simultaneous mass pore production. Thus, not only can TCLB drive novel nanopore sensing applications, TCLB can simultaneously drive the industrial scaling of these applications. As an example, consider combining TCLB with photothermally assisted thinning.[27,42,48] In a photothermally





**Figure 7.** DNA translocation through a nanopore fabricated using TCLB (V<sub>pulse</sub> = 20 V,  $t_{\text{pulse}} = 150$  ms, membrane thickness 10 nm, tip radius *r* = 10 ± 5 nm). a) Typical ionic current traces of DNA translocating through a 9.9 nm pore in a 10 nm thick silicon nitride membrane. The translocation experiment was performed with 0.5 μg mL<sup>−1</sup> 100 bp ladder DNA (100–2000 bp) in 1 м KCl buffered with 10 × 10<sup>−3</sup> м Tris, 1 × 10<sup>−3</sup> м ethylenediaminetetraacetic acid (EDTA), at pH = 8.0. Observed events are labeled as 1-3, corresponding to different DNA configurations/folding states while translocating through the pore. b) Zoomed-in current trace of event 1, 2, and 3, corresponding to the cartoon translocation types shown in (c). d) Current blockade histogram including over 500 events. e) *I*–*V* characterization of the nanopore. The nanopore displays an Ohmic *I*–*V* curve with a resistance of 23.0 MΩ, leading to an effective pore diameter of 9.9 nm. Power spectral density (PSD) of the nanopore is shown in Figure S5 (Supporting Information).

assisted thinning process, a laser beam is focused on a silicon nitride membrane, leading to formation of a locally thinned out region, with thinning achieved down to a few nanometers.[42] If there is only one thinned well formed, classic dielectric breakdown will tend to form a pore at this "thinned out" weakest position. Classic dielectric breakdown, however, is limited to forming only one pore in one well across an entire membrane. In contrast, TCLB can position pores in each member of a large-scale array of photothermally thinned wells, with the wells packed as close as the photothermal thinning technique allows. Specifically, AFM topographic scans will determine the center-point of each well and TCLB will then form pores at these positions.

TCLB may also have applications beyond nanopore fabrication, providing an AFM-based approach to locally characterize the dielectric strength of thin membranes and 2D materials. This application, which could be useful for the microelectromechanical systems (MEMS) and the semiconductor industry, could enable mapping of dielectric strength across large membranes and semiconductor devices, leading to enhanced material performance (e.g., for high-κ gate dielectrics<sup>[49]</sup>).

### **4. Experimental Section**

*Materials*: The silicon nitride membranes used are commercially available from Norcada (part # NBPT005YZ-HR and NT002Y). The membrane was supported by a circular silicon frame (2.7 μm diameter, 200  $\mu$ m thickness) with a window size of 10  $\times$  10, 20  $\times$  20, or  $50 \times 50$   $\mu$ m<sup>2</sup>. The membrane thickness was 10 nm, 12–14 nm, or 20 nm. The AFM probes used were obtained from Adama Innovations (part # AD-2.8-AS) and had a tip radii of curvature of  $10 \pm 5$  nm. Nanopore fabrication experiments were performed in 1 <sup>m</sup> sodium percholorate dissolved in propylene carbonate (PC), with a conductivity of 2.82 S m<sup>-1</sup>.<sup>[50]</sup> DNA translocation experiments were performed in a 3D printed fluidic **www.advancedsciencenews.com www.small-methods.com**

cell with 100 bp ladder DNA (Sigma-Aldrich, 100–2000 bp) diluted to a

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final concentration of 0.5  $\mu$ g mL $^{-1}$  in 1 м KCl buffered with 10  $\times$  10 $^{-3}$  м Tris and  $1 \times 10^{-3}$  m EDTA at pH = 8.0. *Instrumentation*: The atomic force microscope used in our experiments

was a MultiMode Nanoscope III from Digital Instruments (now Bruker). Nanoscript was used for automated fabrication of nanopores. The TEM images were acquired using the JEM-2100F TEM from JEOL.

*Current Data Acquisition and Analysis*: The current signal during nanopore fabrication was recorded using a custom current amplifier with 5 kHz detection bandwidth at the sampling rate of 5k samples per second. A current limiting resistor (2 GΩ) was connected in series with the membrane during the pore fabrication process. Analysis of dielectric breakdown events in the current signal was performed using a custom Python code. The ionic transpore current during DNA translocations was recorded using an Axopatch 200B with a 250 kHz sampling rate, lowpass filtered at 100 kHz. DNA translocation data analysis was carried out using Transalyzer.[51]

## **Supporting Information**

Supporting Information is available from the Wiley Online Library or from the author.

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## **Conflict of Interest**

The authors declare no conflict of interest.

### **Keywords**

AFM, dielectric breakdown, nanopores, single molecule sensing, tip-controlled local breakdown (TCLB)

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